**Cell Line and Biological Materials Pathogen Testing**

When biological material is introduced into an animal, it is a potential source of contamination by pathogens if the material originated from an infected animal, or if it was maintained in an incubator along with an infected source. Pathogens present in these cell lines have the potential to infect animal that are inoculated with them. PCR-based testing of biologic materials is available. In order to protect the animal colonies at TTUHSC, the IACUC requires that all biologics of animal or human origin be tested.

The goal is that no animals should be exposed to human or animal biological materials before the materials have undergone PCR testing and have been verified pathogen-free.

As this is a new program, current research protocols need not stop or be held up, but all biologic materials being used should be screened. Cell lines re-animated from frozen must be tested before use in animals.

Testing requires samples from the investigative staff and takes about 2 weeks from submission for results to come back. Submissions will occur in batches to minimize shipping costs and will be done through the LARC.

**What should be routinely tested?**

* Rodent derived cell lines, transplantable tumors, serum, tissues, body fluids, antibody preparations, and hybridomas to be used in rodents.
  + If the biological material is from a donor animal in the same facility and same room as the recipient, no testing is required.
* Non-rodent derived cell lines, transplantable tumors, serum, tissues, body fluids, antibody preparations, and hybridomas that have been passaged through rodents or have been exposed to rodents outside LARC animal facilities.
* All human origin biologics to be used in animals.
* Biologics of commercial origin can be exempted from testing if the vendor can provide a copy of negative PCR pathogen screening results to LARC.

**Requirements for submission**

* Two 1ml cryovial of each cell line sample containing a minimum of 2 x 106 cells.
  + Cells can be in pellet form, in freeze media, growth media, or PBS.
* For liquid samples two 1ml cryovials with at least 0.5ml.
  + For antibody preparations the volume can be smaller (please contact us).
* Samples should be collected aseptically.
* Samples should be appropriately labeled with Principle Investigator name and cell line designation.
* Completed submission form.

Please complete the information below:

|  |  |
| --- | --- |
| Principal Investigator | Laboratory Contact |
|  |  |
| Email | Email |
|  |  |
| Phone | Phone |
|  |  |

**Samples submitted**

|  |  |  |  |
| --- | --- | --- | --- |
| **Name** | | **Description / Origin** | **Media** |
| Example: Hep93C | | Human hepatocellular carcinoma line | PBS |
| 1. |  |  |  |
| 2. |  |  |  |
| 3. |  |  |  |
| 4. |  |  |  |
| 5. |  |  |  |
| 6. |  |  |  |
| 7. |  |  |  |
| 8. |  |  |  |
| 9. |  |  |  |
| 10. |  |  |  |
| 11. |  |  |  |
| 12. |  |  |  |

\*Attach additional sheets if necessary

Would you like cell line authentication as well (no additional sample needed)?

YES

NO

|  |
| --- |
| Additional comments: |
|  |

* Contact a LARC Vet Tech for submission of your samples.
* Results will be forwarded to the submitter and maintained by LARC.
* Questions regarding submissions can be directed to Dr. Scott Trasti ([scott.trasti@ttuhsc.edu](mailto:scott.trasti@ttuhsc.edu)).

**Appendix I (Excluded Pathogens): Idexx Impact 3 profile.**

* Mycoplasma spp.
* *Mycoplasma pulmonis*
* Mouse Hepatitis Virus
* Minute Virus of Mice
* Mouse Parvovirus
* Theiler’s murine encephalomyelitis virus
* Sendai virus
* Pneumonia Virus of Mice
* Murine Norovirus
* Reovirus 3
* Mouse Rotavirus
* Ectromelia Virus
* Lymphocytic Choriomeningitis Virus
* Polyomavirus
* Lactate dehydrogenase-elevating Virus
* Mouse Kidney Parvovirus\*

\*immunodeficient animals only